Package ‘condvis’

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R topics documented:

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condvis-package

Description

Exploring statistical models by interactively taking 2-D and 3-D sections in data space. The main functions for end users are `ceplot` (see example below) and `condtour`. Requires XQuartz on Mac OS, and X11 on Linux. A website for the package is available at markajoc.github.io/condvis. Source code is available to browse at GitHub. Bug reports and feature requests are very welcome at GitHub.

Details

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arrangeC

References


Examples

```r
## Not run:
mtcars$cyl <- as.factor(mtcars$cyl)
mtcars$am <- as.factor(mtcars$am)

library(mgcv)
model1 <- list(
quadratic = lm(mpg ~ cyl + am + qsec + wt + I(wt^2), data = mtcars),
    additive = gam(mpg ~ cyl + am + qsec + s(wt), data = mtcars))

ceplot(data = mtcars, model = model1, sectionvars = "wt")

## End(Not run)
```

arrangeC

*Make a list of variable pairings for condition selecting plots produced by plotxc*

Description

This function arranges a number of variables in pairs, ordered by their bivariate relationships. The goal is to discover which variable pairings are most helpful in avoiding extrapolations when exploring the data space. Variable pairs with strong bivariate dependencies (not necessarily linear) are chosen first. The bivariate dependency is measured using savingby2d. Each variable appears in the output only once.

Usage

```r
arrangeC(data, method = "default")
```

Arguments

- **data**: A dataframe
- **method**: The character name for the method to use for measuring bivariate dependency, passed to savingby2d.

Details

If `data` is so big as to make `arrangeC` very slow, a random sample of rows is used instead. The bivariate dependency measures are rough, and the ordering algorithm is a simple greedy one, so it is not worth allowing it too much time. This function exists mainly to provide a helpful default ordering/pairing for `ceplot`.
Value

A list containing character vectors giving variable pairings.

References


See Also

savingby2d

Examples

data(powerplant)

pairings <- arrangeC(powerplant)

dev.new(height = 2, width = 2 * length(pairings))
par(mfrow = c(1, length(pairings)))

for (i in seq_along(pairings)){
  plotxc(powerplant[, pairings[[i]]], powerplant[1, pairings[[i]]],
    select.col = NA)
}

---

**ceplot**

*Interactive conditional expectation plot*

Description

Creates an interactive conditional expectation plot, which consists of two main parts. One part is a single plot depicting a section through a fitted model surface, or conditional expectation. The other part shows small data summaries which give the current condition, which can be altered by clicking with the mouse.

Usage

ceplot(data, model, response = NULL, sectionvars = NULL,
conditionvars = NULL, threshold = NULL, lambda = NULL,
distance = c("euclidean", "maxnorm"), type = c("default", "separate",
"shiny"), view3d = FALSE, Corder = "default", selectortype = "minimal",
conf = FALSE, probs = FALSE, col = "black", pch = NULL,
residuals = FALSE, xsplotpar = NULL, modelpar = NULL,
xplotpar = NULL)
Arguments

data A dataframe containing the data to plot
model A model object, or list of model objects
response Character name of response in data
sectionvars Character name of variable(s) from data on which to take a section, can be of length 1 or 2.
conditionvars Character names of conditioning variables from data. These are the predictors which we can set to single values in order to produce a section. Can be a list of vectors of length 1 or 2. Can be a character vector, which is then paired up using arrangeC. If NULL, an attempt will be made to extract all variable names which are not response or sectionvars from model, and these will be arranged using arrangeC.
threshold This is a threshold distance. Points further than threshold away from the current section will not be visible. Passed to similarityweight.
lambda A constant to multiply by number of factor mismatches in constructing a general dissimilarity measure. If left NULL, behaves as though lambda is set greater than threshold, and so only observations whose factor levels match the current section are visible. Passed to similarityweight.
distance A character vector describing the type of distance measure to use, either "euclidean" (default) or "maxnorm".
type This specifies the type of interactive plot. "default" places everything on one device. "separate" places condition selectors on one device and the section on another. (These two options require XQuartz on OS X). "shiny" produces a Shiny application.
view3d Logical; if TRUE plots a three-dimensional regression surface if possible.
corder Character name for method of ordering conditioning variables. See arrangeC.
selectortype Type of condition selector plots to use. Must be "minimal" if type is "default". If type is "separate", can be "pcp" (see plotxc.pcp) or "full" (see plotxc.full).
conf Logical; if TRUE plots confidence bounds (or equivalent) for models which provide this.
probs Logical; if TRUE, shows predicted class probabilities instead of just predicted classes. Only available if S specifies two numeric predictors and the model's predict method provides this.
col Colour for observed data.
pch Plot symbols for observed data.
residuals Logical; if TRUE, plots a residual versus predictor plot instead of the usual scale of raw response.
xsplotpar Plotting parameters for section visualisation as a list, passed to plotxs. Can specify xlim, ylim.
modelpar Plotting parameters for models as a list, passed to plotxs. Not used.
xcplotpar Plotting parameters for condition selector plots as a list, passed to plotxc. Can specify col for highlighting current section, cex, and trim (see plotxc).
References


See Also

condtour, similarityweight

Examples

```r
## Not run:
## Example 1: Multivariate regression, xs one continuous predictor

mtcars$cyl <- as.factor(mtcars$cyl)
library(mgcv)
model1 <- list(
  quadratic = lm(mpg ~ cyl + hp + wt + I(wt^2), data = mtcars),
  additive = mgcv::gam(mpg ~ cyl + hp + s(wt), data = mtcars))
conditionvars1 <- list(c("cyl","hp"))
ceplot(data = mtcars, model = model1, response = "mpg", sectionvars = "wt", conditionvars = conditionvars1, threshold = 0.3, conf = T)

## Example 2: Binary classification, xs one categorical predictor

mtcars$cyl <- as.factor(mtcars$cyl)
library(e1071)
model2 <- list(
  svm = svm(am ~ mpg + wt + cyl, data = mtcars, family = "binomial"),
  glm = glm(am ~ mpg + wt + cyl, data = mtcars, family = "binomial"))
ceplot(data = mtcars, model = model2, sectionvars = "wt", threshold = 1, type = "shiny")

## Example 3: Multivariate regression, xs both continuous

mtcars$cyl <- as.factor(mtcars$cyl)
mtcars$gear <- as.factor(mtcars$gear)
library(e1071)
model3 <- list(svm(mpg ~ wt + qsec + cyl + hp + gear, data = mtcars, family = "binomial"))
conditionvars3 <- list(c("cyl","gear"), "hp")
ceplot(data = mtcars, model = model3, sectionvars = c("wt", "qsec"), threshold = 1, conditionvars = conditionvars3)
```
condtour

condtour(data = mtcars, model = model3, sectionvars = c("wt", "qsec"),
          threshold = 1, type = "separate", view3d = T)

## Example 4: Multi-class classification, xs both categorical

mtcars$cyl <- as.factor(mtcars$cyl)
mtcars$vs <- as.factor(mtcars$vs)
mtcars$am <- as.factor(mtcars$am)
mtcars$gear <- as.factor(mtcars$gear)
mtcars$carb <- as.factor(mtcars$carb)

library(e1071)
model4 <- list(svm(carb ~ ., data = mtcars, family = "binomial"))
ceplot(data = mtcars, model = model4, sectionvars = c("cyl", "gear"),
       threshold = 3)

## Example 5: Multi-class classification, xs both continuous

data(wine)
wine$Class <- as.factor(wine$Class)
library(e1071)

model5 <- list(svm(Class ~ ., data = wine, probability = TRUE))
ceplot(data = wine, model = model5, sectionvars = c("Hue", "Flavanoids"),
       threshold = 3, type = "separate")
ceplot(data = wine, model = model5, sectionvars = c("Hue", "Flavanoids"),
       threshold = 3, type = "separate")
ceplot(data = wine, model = model5, sectionvars = c("Hue", "Flavanoids"),
       threshold = 3, type = "separate", selectortype = "pcc")

## Example 6: Multi-class classification, xs with one categorical predictor,
## and one continuous predictor.

mtcars$cyl <- as.factor(mtcars$cyl)
mtcars$carb <- as.factor(mtcars$carb)

library(e1071)
model6 <- list(svm(cyl ~ carb + wt + hp, data = mtcars, family = "binomial"))
ceplot(data = mtcars, model = model6, threshold = 1, sectionvars = c("carb",
                                                            "wt"), conditionvars = "hp")

## End(Not run)
Description

Whereas `ceplot` allows the user to interactively choose sections to visualise, `condtour` allows the user to pre-select all sections to visualise, order them, and cycle through them one by one. `']' key advances the tour, and `[' key goes back. Can adjust threshold for the current section visualisation with `',' and `.' keys.

Usage

```r
condtour(data, model, path, response = NULL, sectionvars = NULL,
conditionvars = NULL, threshold = NULL, lambda = NULL,
distance = c("euclidean", "maxnorm"), view3d = FALSE,
Corder = "default", conf = FALSE, col = "black", pch = NULL,
xplotpar = NULL, modelpar = NULL, xcplotpar = NULL)
```

Arguments

- `data` A dataframe.
- `model` A fitted model object, or a list of such objects.
- `path` A dataframe, describing the sections to take. Basically a dataframe with its colnames being `conditionvars`.
- `response` Character name of response variable in `data`.
- `sectionvars` Character name(s) of variables in `data` on which to take sections.
- `conditionvars` Character name(s) of variables in `data` on which to condition.
- `threshold` Threshold distance. Observed data which are a distance greater than `threshold` from the current section are not visible. Passed to `similarityweight`.
- `lambda` A constant to multiply by number of factor mismatches in constructing a general dissimilarity measure. If left `NULL`, behaves as though `lambda` is set greater than `threshold`, and so only observations whose factor levels match the current section are visible. Passed to `similarityweight`.
- `distance` The type of distance measure to use, either "euclidean" (default) or "maxnorm".
- `view3d` Logical; if `TRUE`, plots a three-dimensional regression surface when possible.
- `Corder` Character name for method of ordering conditioning variables. See `arrangeC`.
- `conf` Logical; if `TRUE`, plots confidence bounds or equivalent when possible.
- `col` Colour for observed data points.
- `pch` Plot symbols for observed data points.
- `xplotpar` Plotting parameters for section visualisation as a list, passed to `plotx`. Not used.
- `modelpar` Plotting parameters for models as a list, passed to `plotx`. Not used.
- `xcplotpar` Plotting parameters for condition selector plots as a list, passed to `plotxc`. Can specify `cex.axis, cex.lab, tck, col` for highlighting current section, `cex`.
Value

Produces a set of interactive plots. One device displays the current section. A second device shows the current section in the space of the conditioning predictors given by `conditionvars`. A third device shows some simple diagnostic plots; one to show approximately how much data are visible on each section, and another to show what proportion of data are visited by the tour.

See Also

`ceplot`, `similarityweight`

Examples

```r
## Not run:

data(powerplant)
library(e1071)
model <- svm(PE ~ ., data = powerplant)
path <- makepath(powerplant[-5], 25)
condtour(data = powerplant, model = model, path = path$path,
          sectionvars = "AT")

data(wine)
wine$class <- as.factor(wine$Class)
library(e1071)
model5 <- list(svm(Class ~ ., data = wine))
conditionvars1 <- setdiff(colnames(wine), c("Class", "Hue", "Flavanoids"))
path <- makepath(wine[, conditionvars1], 50)
condtour(data = wine, model = model5, path = path$path, sectionvars = c("Hue"
                   , "Flavanoids"), threshold = 3)

## End(Not run)
```

---

**cont2color** Assign colours to numeric vector

Description

This function assigns colours on a linear scale to a numeric vector. Default is to try to use RColorBrewer for colours, and `cm.colors` otherwise. Can provide custom range, breaks and colours.

Usage

```r
cont2color(x, xrange = NULL, breaks = NULL, colors = NULL)
```
**Arguments**

- **x**: A numeric vector.
- **xrange**: The range to use for the colour scale.
- **breaks**: The number of breaks at which to change colour.
- **colors**: The colours to use. Defaults to a diverging colour scheme; either "PiYG" from RColorBrewer if available, or cm.colors otherwise.

**Details**

Uses the RColorBrewer package if installed. Coerces x to numeric with a warning.

**Value**

A character vector of colours.

**See Also**

factor2color

**Examples**

```r
x <- runif(200)
plot(x, col = cont2color(x, c(0,1)))
plot(x, col = cont2color(x, c(0,0.5)))
plot(sort(x), col = cont2color(sort(x), c(0.25,0.75)), pch = 16)
abline(h = c(0.25, 0.75), lty = 3)
```

---

**crab**

*Brockmann's crab data*

**Description**

Abstract from original paper: Horseshoe crabs arrive on the beach in pairs and spawn in the high intertidal during the springtime, new and full moon high tides. Unattached males also come to the beach, crowd around the nesting couples and compete with attached males for fertilizations. Satellite males form large groups around some couples while ignoring others, resulting in a non-random distribution that cannot be explained by local environmental conditions or habitat selection. In experimental manipulations, pairs that had satellites regained them after they had been removed whereas pairs with no satellites continued nesting alone, which means that satellites were not simply accumulating around the pairs that had been on the beach the longest. Manipulations also revealed that satellites were not just copying the behaviour of other males. Based on the evidence from observations and experiments, the most likely explanation for the nonrandom distribution of satellite males among nesting pairs is that unattached males are preferentially attracted to some females over others. Females with many satellites were larger and in better condition, but did not lay more eggs,
than females with few or no satellites.

- satellites: response variable; number of satellites around female crab
- color: color of crab
- spine: condition of spine
- weight: weight of crab
- width: width of carapace

**Format**

173 observations on 5 variables.

**Source**

https://onlinecourses.science.psu.edu/stat504/node/169

**References**


**Examples**

```r
data(crab)
```

**Description**

Calculate Minkowski distance between one point and a set of other points.

**Usage**

```r
dist1(x, X, p = 2, inf = FALSE)
```

**Arguments**

- `x`: A numeric vector describing point coordinates.
- `X`: A numeric matrix describing coordinates for several points.
- `p`: The power in Minkowski distance, defaults to 2 for Euclidean distance.
- `inf`: Logical; switch for calculating maximum norm distance (sometimes known as Chebyshev distance) which is the limit of Minkowski distance as `p` tends to infinity.

**Value**

A numeric vector. These are `distance^p`, for speed of computation.
factor2color

Assign colours to factor vector

Description

This function takes a factor vector and returns suitable colours representing the factor levels. Default is to try to use RColorBrewer for colours, and rainbow otherwise. Can provide custom colours.

Usage

factor2color(x, colors = NULL)

Arguments

x

A factor vector.

colors

The colours to use. Defaults to a qualitative colour scheme; either "Set3" from RColorBrewer if available, or rainbow otherwise.

Details

Uses the RColorBrewer package if installed. Coerces x to factor with a warning.

Value

A character vector of colours.

See Also

similarityweight

Examples

x <- runif(5000)
y <- runif(5000)
x1 <- 0.5
y1 <- 0.5

dev.new(width = 4, height = 5.3)
par(mfrow = c(2, 2))

for(p in c(0.5, 1, 2, 10)){
  d <- dist1(x = c(x1, y1), X = cbind(x, y), p = p) ^ (1/p)
col <- rep("black", length(x))
col[d < 0.3] <- "red"
  plot(x, y, pch = 16, col = col, asp = 1, main = paste("p = ", p, sep = ""))
}

---

factor2color

Assign colours to factor vector

Description

This function takes a factor vector and returns suitable colours representing the factor levels. Default is to try to use RColorBrewer for colours, and rainbow otherwise. Can provide custom colours.

Usage

factor2color(x, colors = NULL)

Arguments

x

A factor vector.

colors

The colours to use. Defaults to a qualitative colour scheme; either "Set3" from RColorBrewer if available, or rainbow otherwise.

Details

Uses the RColorBrewer package if installed. Coerces x to factor with a warning.

Value

A character vector of colours.

See Also

similarityweight

Examples

x <- runif(5000)
y <- runif(5000)
x1 <- 0.5
y1 <- 0.5

dev.new(width = 4, height = 5.3)
par(mfrow = c(2, 2))

for(p in c(0.5, 1, 2, 10)){
  d <- dist1(x = c(x1, y1), X = cbind(x, y), p = p) ^ (1/p)
col <- rep("black", length(x))
col[d < 0.3] <- "red"
  plot(x, y, pch = 16, col = col, asp = 1, main = paste("p = ", p, sep = ""))
}
Interpolate a numeric or factor vector.

Usage

\[ \text{interpolate}(x, \ldots) \]

## S3 method for class 'numeric'
\[ \text{interpolate}(x, \text{ninterp} = 4L, \ldots) \]

## S3 method for class 'integer'
\[ \text{interpolate}(x, \text{ninterp} = 4L, \ldots) \]

## S3 method for class 'factor'
\[ \text{interpolate}(x, \text{ninterp} = 4L, \ldots) \]

## S3 method for class 'character'
\[ \text{interpolate}(x, \text{ninterp} = 4L, \ldots) \]

Arguments

- \( x \): A numeric or factor vector.
- \( \ldots \): Not used.
- \( \text{ninterp} \): The number of points to interpolate between observations. It should be an even number for sensible results on a factor/character vector.
makepath

Make a default path for conditional tour

Description

Provides a default path (a set of sections), useful as input to a conditional tour (condtour). Clusters the data using k-means or partitioning around medoids (from the cluster package). The cluster centres/prototypes are then ordered to create a sensible way to visit each section as smoothly as possible. Ordering uses either the DendSer or TSP package. Linear interpolation is then used to create intermediate points between the path nodes.

Usage

makepath(x, ncentroids, ninterp = 4)

Arguments

x A dataframe
ncentroids The number of centroids to use as path nodes.
ninterp The number of points to linearly interpolate between path nodes.

Value

A list with two dataframes: centers giving the path nodes, and path giving the full interpolated path.

See Also

condtour

Examples

d <- data.frame(x = runif(500), y = runif(500))
plot(d)
mp1 <- makepath(d, 5)
points(mp1$centers, type = "b", col = "blue", pch = 16)
mp2 <- makepath(d, 40)
points(mp2$centers, type = "b", col = "red", pch = 16)
plotxc

Condition selector plot

Description

Data visualisations used to select sections for ceplot.

Usage

plotxc(xc, xc.cond, name = NULL, trim = NULL, select.colour = NULL, select.lwd = NULL, cex.axis = NULL, cex.lab = NULL, tck = NULL, select.cex = 1, hist2d = NULL, fullbin = NULL, ...)

Arguments

xc A numeric or factor vector, or a dataframe with two columns
xc.cond Same type as xc, representing a single point in data space to highlight.
name The variable name for xc
trim Logical; if TRUE, long tails of continuous data are chopped off at the 5th and 95th percentiles.
select.colour Colour to highlight xc.cond
select.lwd Line weight to highlight xc.cond
cex.axis Axis text scaling
cex.lab Label text scaling
tck Plot axis tick size
select.cex Plot symbol size
hist2d If TRUE, a scatterplot is visualised as a 2-D histogram. Default behaviour is to use a 2-D histogram if there are over 2,000 observations.
fullbin A cap on the counts in a bin for the 2-D histogram, helpful with skewed data. Larger values give more detail about data density. Defaults to 25.
... Passed to condvis:::spineplot2.

Value

Produces a plot, and returns a list containing the relevant information to update the plot at a later stage.

References

See Also

- `ceplot.plotx`
- `plotx, ceplot, condtour`

Examples

```r
## Histogram, highlighting the first case.

data(mtcars)
obj <- plotxc(mtcars[, "mpg"], mtcars[1, "mpg"])
obj$usr

## Barplot, highlighting 'cyl' = 6.
plotxc(as.factor(mtcars[, "cyl"])), 6, select.colour = "blue")

## Scatterplot, highlighting case 25.
plotxc(mtcars[, c("qsec", "wt")], mtcars[25, c("qsec", "wt")],
select.colour = "blue", select.lwd = 1, lty = 3)

## Boxplot, where 'xc' contains one factor, and one numeric.
mtcars$carb <- as.factor(mtcars$carb)
plotxc(mtcars[, c("carb", "wt")], mtcars[25, c("carb", "wt")],
select.colour = "red", select.lwd = 3)

## Spineplot, where 'xc' contains two factors.
mtcars$gear <- as.factor(mtcars$gear)
mtcars$cyl <- as.factor(mtcars$cyl)
plotxc(mtcars[, c("cyl", "gear")], mtcars[25, c("cyl", "gear")],
select.colour = "red")

## Effect of 'trim'.

x <- c(-200, runif(400), 200)
plotxc(x, 0.5, trim = FALSE, select.colour = "red")
plotxc(x, 0.5, trim = TRUE, select.colour = "red")
```

plotxc.pcp  

Condition selector plot

Description

Multivariate data visualisations used to select sections for `ceplot`. Basically visualises a dataset and highlights a single point.
plotxs

Usage

plotxc.pcp(Xc, Xc.cond, select.colour = NULL, select.lwd = 3, cex.axis = NULL, cex.lab = NULL, tck = NULL, select.cex = 1, ...)

plotxc.full(Xc, Xc.cond, select.colour = NULL, select.lwd = 3, cex.axis = NULL, cex.lab = NULL, tck = NULL, select.cex = 0.6, ...)

Arguments

Xc A dataframe.
Xc.cond A dataframe with one row and same names as Xc.
select.colour Colour to highlight Xc.cond
select.lwd Line weight to highlight Xc.cond
cex.axis Axis text scaling
cex.lab Label text scaling	
tck Plot axis tick size
select.cex Plot symbol size
... not used.

Value

Produces a plot, and returns a list containing the relevant information to update the plot at a later stage.

See Also

ceplot, plotxs, plotxc

plotxs Visualise a section in data space

Description

Visualise a section in data space, showing fitted models where they intersect the section, and nearby observations. The weights for observations can be calculated with similarityweight. This function is mainly for use in ceplot and condtour.

Usage

plotxs(xs, y, Xc.cond, model, model.colour = NULL, model.lwd = NULL, model.lty = NULL, model.name = NULL, yhat = NULL, mar = NULL, col = "black", weights = NULL, view3d = FALSE, theta3d = 45, phi3d = 20, xs.grid = NULL, prednew = NULL, conf = FALSE, probs = FALSE, pch = 1, residuals = FALSE, main = NULL, xlim = NULL, ylim = NULL)
Arguments

- **xs**: A dataframe with one or two columns.
- **y**: A dataframe with one column.
- **xc.cond**: A dataframe with a single row, with all columns required for passing to `predict` methods of models in `model`.
- **model**: A fitted model object, or a list of such objects.
- **model.colour**: Colours for fitted models. If `model` is a list, this should be of same length as `model`.
- **model.lwd**: Line weight for fitted models. If `model` is a list, this should be of same length as `model`.
- **model.lty**: Line style for fitted models. If `model` is a list, this should be of same length as `model`.
- **model.name**: Character labels for models, for legend.
- **yhat**: Fitted values for the observations in `y`. Calculated if needed and not provided. Only used if showing residuals, or `xs` has two columns.
- **mar**: Margins for plot.
- **col**: Colours for observed data. Should be of length `nrow(xs)`.
- **weights**: Similarity weights for observed data. Should be of length `nrow(xs)`. Usually calculated with `similarityweight`.
- **view3d**: Logical; if TRUE plots a three-dimensional regression surface if possible.
- **theta3d, phi3d**: Angles defining the viewing direction. theta3d gives the azimuthal direction and phi3d the colatitude. See `persp`.
- **xs.grid**: The grid of values defining the part of the section to visualise. Calculated if not provided.
- **prednew**: The y values where the models in `model` intersect the section. Useful when providing theta3d, phi3d, or weights, where the predict methods have been called elsewhere.
- **conf**: Logical; if TRUE plots confidence bounds (or equivalent) for models which provide this.
- **probs**: Logical; if TRUE, shows predicted class probabilities instead of just predicted classes. Only available if `xs` contains two numeric predictors and the model’s predict method provides this.
- **pch**: Plot symbols for observed data.
- **residuals**: Logical; if TRUE, plots a residual versus predictor plot instead of the usual scale of raw response.
- **main**: Character title for plot, default is "Conditional expectation".
- **xlim**: Graphical parameter passed to plotting functions.
- **ylim**: Graphical parameter passed to plotting functions.

Value

A list containing relevant information for updating the plot.
powerplant

References


See Also

plotxc, ceplot, condtour

Examples

data(mtcars)
model <- lm(mpg ~ ., data = mtcars)
plotxs(xs = mtcars[, "wt", drop = FALSE], y = mtcars[, "mpg", drop = FALSE],
     xc.cond = mtcars[1, ], model = list(model))

powerplant  Tuefekci’s powerplant data

Description

The dataset contains 9568 data points collected from a Combined Cycle Power Plant over 6 years (2006-2011), when the power plant was set to work with full load. Features consist of hourly average ambient variables Temperature (T), Ambient Pressure (AP), Relative Humidity (RH) and Exhaust Vacuum (V) to predict the net hourly electrical energy output (EP) of the plant.

A combined cycle power plant (CCPP) is composed of gas turbines (GT), steam turbines (ST) and heat recovery steam generators. In a CCPP, the electricity is generated by gas and steam turbines, which are combined in one cycle, and is transferred from one turbine to another. While the Vacuum is collected from and has effect on the Steam Turbine, the other three of the ambient variables affect the GT performance.

Format

9568 observations on 5 continuous variables.

Source


References

Examples

data(powerplant)
head(powerplant)

savingby2d Assess advantage of 2-D view over 1-D view for identifying extrapolation

Description
A simple algorithm to evaluate the advantage of by taking a bivariate marginal view of two variables, when trying to avoid extrapolations, rather than two univariate marginal views.

Usage
savingby2d(x, y = NULL, method = "default")

Arguments

x A numeric or factor vector. Can also be a dataframe containing x and y, if y is NULL.
y A numeric or factor vector.
method Character; criterion used to quantify bivariate relationships. Can be "default", a scagnostic measure, or "DECR" to use a density estimate confidence region.

Details
If given two continuous variables, the variables are both scaled to mean 0 and variance 1. Then the returned value is the ratio of the area of the convex hull of the data to the area obtained from the product of the ranges of the two areas, i.e. the area of the bounding rectangle.
If given two categorical variables, all combinations are tabulated. The returned value is the number of non-zero table entries divided by the total number of table entries.
If given one categorical and one continuous variable, the returned value is the weighted mean of the range of the continuous variable within each category divided by the overall range of the continuous variable, where the weights are given by the number of observations in each level of the categorical variable.
Requires package scagnostics if a scagnostics measure is specified in method. Requires package hdrcde if "DECR" (density estimate confidence region) is specified in method. These only apply to cases where x and y are both numeric.

Value
A number between 0 and 1. Values near 1 imply no benefit to using a 2-D view, whereas values near 0 imply that a 2-D view reveals structure hidden in the 1-D views.
similarityweight

References


See Also

\texttt{similarityweight}

Examples

\begin{verbatim}
x <- runif(1000)
y <- runif(1000)
plot(x, y)
savingby2d(x, y)
## value near 1, no real benefit from bivariate view

x1 <- runif(1000)
y1 <- x1 + rnorm(sd = 0.3, n = 1000)
plot(x1, y1)
savingby2d(x1, y1)
## smaller value indicates that the bivariate view reveals some structure
\end{verbatim}

---

\texttt{similarityweight} \hspace{1cm} \textit{Calculate the similarity weight for a set of observations}

Description

Calculate the similarity weight for a set of observations, based on their distance from some arbitrary points in data space. Observations which are very similar to the point under consideration are given weight 1, while observations which are dissimilar to the point are given weight zero.

Usage

\texttt{similarityweight(x, data, threshold = NULL, distance = NULL, lambda = NULL)}

Arguments

\begin{itemize}
  \item \textbf{x} \hspace{1cm} A dataframe describing arbitrary points in the space of the data (i.e., with same colnames as data).
  \item \textbf{data} \hspace{1cm} A dataframe representing observed data.
  \item \textbf{threshold} \hspace{1cm} Threshold distance outside which observations will be assigned similarity weight zero. This is numeric and should be > 0. Defaults to 1.
  \item \textbf{distance} \hspace{1cm} The type of distance measure to be used, currently just two types of Minkowski distance: "euclidean" (default), and "maxnorm".
\end{itemize}
A constant to multiply by the number of categorical mismatches, before adding to the Minkowski distance, to give a general dissimilarity measure. If left NULL, behaves as though lambda is set larger than threshold, meaning that one factor mismatch guarantees zero weight.

Details

Similarity weight is assigned to observations based on their distance from a given point. The distance is calculated as Minkowski distance between the numeric elements for the observations whose categorical elements match, with the option to use a more general dissimilarity measure comprising Minkowski distance and a mismatch count.

Value

A numeric vector or matrix, with values from 0 to 1. The similarity weights for the observations in data arranged in rows for each row in x.

References


See Also
dist1

Examples

```r
## Say we want to find observations similar to the first observation.
## The first observation is identical to itself, so it gets weight 1. The
## second observation is similar, so it gets some weight. The rest are more
## different, and so get zero weight.

data(mtcars)
similarityweight(x = mtcars[1, ], data = mtcars)

## By increasing the threshold, we can find observations which are more
## approximately similar to the first row. Note that the second observation
## now has weight 1, so we lose some ability to discern how similar
## observations are by increasing the threshold.

similarityweight(x = mtcars[1, ], data = mtcars, threshold = 5)

## Can provide a number of points to 'x'. Here we see that the Mazda RX4 Wag
## is more similar to the Merc 280 than the Mazda RX4 is.

similarityweight(mtcars[1:2, ], mtcars, threshold = 3)
```
### Italian wine data

**Description**

<table>
<thead>
<tr>
<th>Class</th>
<th>Alcohol</th>
<th>Malic acid</th>
<th>Ash</th>
<th>Alcalinity</th>
<th>Magnesium</th>
<th>Phenols</th>
<th>Flavanoids</th>
<th>Nonflavanoid</th>
<th>Proanthocyanins</th>
<th>Intensity</th>
<th>Hue</th>
<th>OD280/OD315</th>
<th>Proline</th>
</tr>
</thead>
<tbody>
<tr>
<td>3 different</td>
<td>Alcohol</td>
<td>Malic acid</td>
<td>Ash</td>
<td>Alcalinity</td>
<td>Magnesium</td>
<td>Phenols</td>
<td>Flavanoids</td>
<td>Nonflavanoid</td>
<td>Proanthocyanins</td>
<td>Intensity</td>
<td>Hue</td>
<td>OD280/OD315</td>
<td>Proline</td>
</tr>
<tr>
<td>cultivars</td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Format**

178 observations on 14 variables.

**Source**

UCI repository. [https://archive.ics.uci.edu/ml/datasets/Wine](https://archive.ics.uci.edu/ml/datasets/Wine)

**References**


**Examples**

```r
data(wine)
pairs(wine[, -1], col = factor2color(wine$Class), cex = 0.2)
```
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